ISSN 2278 – 5221 www.ijpmbs.com Vol. 2, No. 2, April 2013 © 2013 IJPMBS. All Rights Reserved

Research Paper

OPTIMIZATION OF PHYSICAL CONDITIONS FOR AS(III) BIOSORPTION BY ASPERGILLUS NIGER X₃₀₀

S Ganguly1*

*Corresponding Author: **S Ganguly,** \boxtimes subhadeepgangulyphysiol@rediffmail.com

An experimental study was conducted to optimize different physical conditions for As(III) bio sorption by an As(III) resistant strain *Aspergillus niger* X_{300} . Maximum biosorption was resulted with initial pH , 4.5; temperature , 30 °C; spore density , 7 X 108; shaker speed , 200 rpm; volume of medium , 100 mL ; age of inoculum , 7 days and incubation period , 7 days.

Keywords: Experimental, Optimize, Physical, As (III), Biosorption

INTRODUCTION

Bio-sorption is a metabolically passive process which is dependent on kinetic equilibrium and the composition of sorbent cellular surface chemistry (Loukidou et al., 2003). Adsorbing elements are adsorbed onto the cellular surface. As it is determined by equilibrium, it is greatly influenced by pH, concentration of biomass and other physical conditions that alter the biomass (Murugesan et al., 2006; Mamisahebei et al., Velasque and Dussan, 2009). Among the many abiotic factors (physical conditions) fungal growth is significantly affected by pH and temperature (Roberts and Cambell, 1977). Considering the reviews, my present study was intended to optimize different Physical conditions namely initial pH, temperature, spore density, agitation/aeration, volume of medium, age of inoculum and incubation period one by one to improve the bio sorption of As(III) by

experimentally developed resistant fungus $Aspergillus\ niger\ X_{300}$ (Ganguly, 2013).

MATERIALS AND METHODS

Microorganism: An experimentally developed As(III) resistant strain *Aspergillus niger* X_{300} was used throughout the study (Ganguly, 2013).

Medium Composition Used for Fungal Growth: The growth medium was composed of: glucose, 10%; urea, 0.8%; MgSO₄.7H₂O, 0.03%; KH₂PO₄, 0.1%; K₂HPO₄, 0.1%; CaCl₂, 0.3%.

Medium Composition for Maintenance Medium: Synthetic maintenance medium contains: glucose, 0.1%; urea , 0.6%; MgSO₄.7H₂O, 0.01%; KH₂PO₄, 0.1%; K₂HPO₄, 0.1%; CaCl₂, 0.2% and agar , 4% as solidifying agent.

Estimation of As(III): The concentration of As(III)

Department of Biological Sciences, Sankrail Abhoy Charan High School (HS), Sankrail, Howrah, West Bengal, India.

in the broth was estimated by the method as reported by Cernansky *et al.* (2007).

Estimation of Dry Cell/Spore Weight: Fungal cells /spores were filtered using Whatmen No.1 filter paper and heated at 70 °C until it becomes dry and its weight was estimated by electronic weighing machine (ECELON MS- 2690).

Statistical Analysis: All data were expressed as Mean \pm SEM. Data were analyzed by one way ANOVA followed by Dunett's *post hoc* multiple comparison test considering p<0.05 as significant and p<0.01 as highly significant (using Prism 4.0).

Experiments were carried out in six sets using 250 mL Erlenmeyer Conical flask using 50 mL growth medium, pH 5.0, temperature, 28 °C, spore/cell density 5x10⁷ cells or spores/mL, shaker speed, 150 rpm, age of inoculum, 7 days and incubation period, 7 days.

RESULTS AND DISCUSSION

Tables 1 to 7 showed the optimization of different physical condition for this bio sorption.

From the tables, it can be tentatively concluded that the fungus *Aspergillus niger* X_{300} grew best with pH, 4.5, temperature, 30 °C, spore density, 7 x 10⁸ spores or cells/mL; shaker speed,

Initial pH	Initial concentration of As(III) [mg/L]	Final concentration of As (III) [mg/L]	Dry cell/spore weight (gm/L
7.0	1500	672.7±6.136 **	6.3±0.431 **
6.5	1500	657.8±4.212 **	6.5±0.661 *
6.0	1500	642.3±4.313 **	6.6±0.731 *
5.5	1500	598.7±3.683 *	6.9±0.613 *
5.0 (Control)	1500	566.4±4.683	7.3±0.683
? 4.5	1500	495.8±3.661 *	8.3±0.832 **
4.0	1500	542.4±6.136 *	7.7±0.661

Table 2: Effect of Temparature (°C)				
°C	Initial Concentration of As(III) [mg/L]	Final Concentration of As(III) [mg/L]	Dry Cell/Spore Weight (g/L)	
25	1500	**696.2±3.396	**6.0±0.661	
26	1500	**631.6±4.136	**6.3±0.432	
27	1500	*591.2±6.213	7.0±0.361	
28 (Control)	1500	566.4±4.683	7.3 ± 0.683	
29	1500	**511.2±3.163	*8.1±0.832	
? 30	1500	**476.6±4.163	**8.6±0.881	
31	1500	**496.2±3.683	**8.9±0.763	

Spores/mL	Initial Concentration of As(III) [mg/L]	Final Concentration of As (III) [mg/L]	Dry Cell/Spore Weight (g/L)
3 × 10 ⁶	1500	**521.8±3.961	8.3±0.661
5 × 10 ⁷ (Control)	1500	476.6±4.163	8.6±0.881
6 × 10 ⁸	1500	*432.4±4.613	8.9±0.731
8 × 7 × 10 ⁸	1500	*401.2±3.921	*9.1±0.683
9 × 10 ¹¹	1500	**422.6±3.613	*9.3±0.661

Table 4: Effect of Agitation/Aeration				
Shaker Speed (rpm)	Initial Concentration of As(III) (mg/L)	Final Concentration of As(III) (mg/L)	Dry Cell/Spore Weight (gm/L)	
50	1500	**522.6±6.213	*8.3±0.861	
100	1500	**464.2±4.831	*8.5±0.832	
150 (Control)	1500	401.2±3.921	9.1±0.683	
7200	1500	*372.8±7.613	*9.9±0.661	
250	1500	*591.9±6.136	*9.7±0.862	
Note: Values were expressed as Mean \pm SEM, where n=6; *p<0.05 and **p<0.01, \blacksquare Stands for maximum bio sorption.				

Table 5: Effect of Volume of Medium				
Volume of Medium	Initial Concentration of As(III) (mg/L)	Final Concentration of As(III) (mg/L)	Dry Cell/Spore Weight (gm/L)	
25	1500	*416.3±6.133	*9.3±0.732	
50 (control)	1500	372.8±7.613	9.9±0.661	
75	1500	**333.2±6.113	*10.4±0.763	
2 100	1500	**301.6±8.616	**10.8±0.669	
150	1500	**321.4±4.663	*10.5±0.613	
Note: Values were expressed as Mean ± SEM, where n=6; *p<0.05 and **p<0.01, Stands for maximum bio sorption.				

200 rpm; volume of medium, 100 mL; age of inoculum, 7 days and incubation period, 7 days. Papagianni (2007) reported the optimum pH for *Aspergillus niger* was 4.5, required dissolved oxygen for its maximum growth. Majumder *et al.* (2010)

claimed that a mutant strain *Aspergillus niger* 14/20 grew best with pH 5 at 30 °C. As the biosorption is directly proportional to the fungal growth, maximumAs (III) biosorption was obtained with the above mentioned physical conditions.

Age of Inoculum (days)	Initial Concentration of As(III) (mg/L)	Final Concentration of As(III) (mg/L)	Dry Cell/Spore Weight (g/L)
1	1500	**1332.6±4.316	**1.6±0.161
2	1500	**922.4±6.613	**3.6±0.642
3	1500	**821.2±5.163	**4.2±0.432
4	1500	**527.4±6.111	**8.3±0.331
5	1500	**419.3±5.213	**8.8±0.683
6	1500	**376.4±7.161	**9.9±0.731
2 7(control)	1500	301.6±8.616	**10.8±0.669
8	1500	*316.6±6.316	**10.9±0.661

Table 7: Optimization of Incubation Period					
Incubation Period(Days)	Initial Concentration of As(III) (mg/L)	Final Concentration of As(III) (mg/L)	Dry Cell/Spore Weight (g/L)		
3	1500	1122.4±7.313	2.9±0.613		
4	1500	774.3±6.224	4.6±0.661		
5	1500	521.6±5.313	8.4±0.461		
6	1500	326.2±6.316	11.1±0.663		
7 (control)	1500	301.6±8.616	10.8±0.669		
8	1500	301.6±6.116	10.8±0.591		

Note: Values were expressed as Mean \pm SEM , where n=6; *p<0.05 and **p<0.01, \blacksquare Stands for maximum bio-sorption.

ACKNOWLEDGMENT

I express my sincere gratitude to the department of Chemical Engineering, University of Calcutta, Bose Institute, Kolkata, Indian Institute of Chemical Biology, Kolkata, Department of Food technology and Biochemical Engineering, Jadavpur University for their kind cooperation without which I could not finish the work.

REFERENCES

 Ganguly S (2013), "Selection of Suitable Fungus, Development of Arsenic Resistant Strain and Kinetic Study for As (III) Biosorption", *Ind. J. Appl. Res.*, Press.

- Loukidou M X, Matis K A, Zouboulis A I and Liakopoulou K M (2003), "Removal of As(V) from Waste Waters by Chemically Modified Fungal Biomass", Water Res., Vol. 37, pp. 4544-4552.
- Mamisahebei S, Khaniki Gh R J, Torabian A, Nasseri S and Naddafi K (2007), "Removal of Arsenic From an Aqueous Solution by Pre-treated Waste Tea Fungal Biomass", *J. Environ. Health. Sci. Eng.*, Vol. 4, pp. 85-92.
- 4. Majumder L, Khalil I, Munshi M K, Alam K, Rashid H, Begum R and Alam N (2010),

- "Citric Acid Production by *Aspergillus Niger* Using Molasses and Pumpkin As Substrates", *Eur. J. Biol. Sci.*, Vol. 2, pp. 1-8.
- 5. Papagianni M (2007), "Advances in Citric Acid Fermentation by *Aspergillus niger*: Biochemical Aspects, Membrane Transport and Modelling", *Biotechnol Adv.*, Vol. 25, pp. 244-263.
- 6. Robert D W and Campbell A S (1977), "Stability of Entomopathogenic Fungi", in
- Hostetter D L and Ignoffo C M (Eds.), *Environmental Stability of Microbial insecticides*, Misllaneous Publications of the Entomological Society of America, Vol. 10, pp. 19-67.
- 7. Velasquez L and Dussan J (2009), "Biosorption and Bio accumulation of Heavy Metals on Dead and Living Biomass of *Bacillus sphaericus*", *J. Hazard. Mater*, Vol. 167, pp. 713-716.